


Patologia para Gastroenterologistas e Endoscopistas

Parte 1: Guidelines Internacionais para o trato gastrointestinal

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Guidelines

60th WSAVA Global Veterinary Conference

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In this section:

- Animal Welfare Guidelines
- Gastrointestinal Guidelines**
- Global Dental Guidelines
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- Renal Standardization Guidelines
- Vaccination Guidelines
- WSAVA Global Zoonosis

Gastrointestinal Guidelines

Standardized GI Endoscopy Reporting Forms

The upper and lower GI endoscopy report forms linked below, represent the work of the WSAVA Working Group on GI Histopathology. The group recognized the need to also standardize endoscopic examination and sampling of the GI tract to ensure the highest procedural diagnostic yield. These endoscopy report forms help address this, namely that endoscopic examination is complete and thorough. Ongoing work is looking at the number of mucosal biopsies needed to ensure diagnosis of different lesions as well as the effect of quality of endoscopic biopsy on the likelihood of diagnosis. The GI Standardization Group hope that we will soon have

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Gastrointestinal Guidelines

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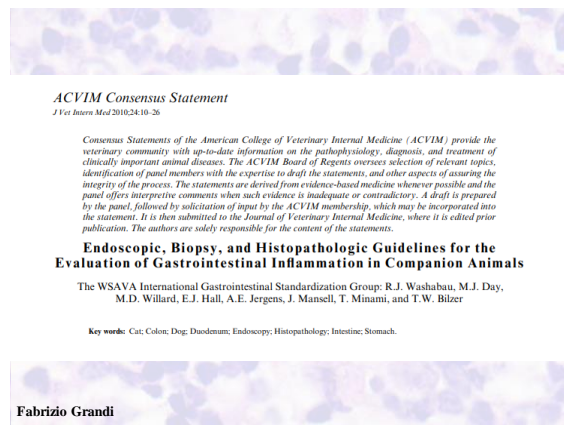
Standardized GI Endoscopy Reporting Forms

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- Upper GI Endoscopy Report Form (PDF).
- Lower GI Endoscopy Report Form (PDF).
- Endoscopic, Biopsy, and Histopathologic Guidelines for the Evaluation of Gastrointestinal Inflammation in Companion Animals (Web).

Endorsement

- These forms have been officially endorsed by the Comparative Gastroenterology Society (CGS) and the European Society of Comparative Gastroenterology (ESCG).



ACVIM Consensus Statement

J Vet Intern Med 2010;24:10-26

Consensus Statements of the American College of Veterinary Internal Medicine (ACVIM) provide the veterinary community with up-to-date information on the pathophysiology, diagnosis, and treatment of clinically important animal diseases. The ACVIM Board of Regents oversees selection of relevant topics, identification of panel members with the expertise to draft the statements, and other aspects of assuring the integrity of the process. The statements are derived from evidence-based medicine wherever possible and the panel offers interpretive comments when such evidence is inadequate or contradictory. A draft is prepared by the panel, followed by solicitation of input by the ACVIM membership, which may be incorporated into the statement. It is then submitted to the *Journal of Veterinary Internal Medicine*, where it is edited prior publication. The authors are solely responsible for the contents of the statements.

Endoscopic, Biopsy, and Histopathologic Guidelines for the Evaluation of Gastrointestinal Inflammation in Companion Animals

The WSAVA International Gastrointestinal Standardization Group: R.J. Washabau, M.J. Day, M.D. Willard, E.J. Hall, A.E. Jergens, J. Mansell, T. Minami, and T.W. Biltzer

Key words: Cat; Colon; Dog; Duodenum; Endoscopy; Histopathology; Intestine; Stomach.

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Considerações prévias à biópsia

- Testes laboratoriais (bioquímico, hemograma)
- Ultrassonografia abdominal
 - Localização da lesão (além do endoscópio)
 - Padrão de lesão
 - Indicação de PAF prévia
 - Contraindicação do exame endoscópico

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The Veterinary Journal

Volume 214, August 2016, Pages 50-60



Review

Maximizing the diagnostic utility of endoscopic biopsy in dogs and cats with gastrointestinal disease

Albert E. Jergens ^a, Michael D. Willard ^b, Karin Allenspach ^c

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<https://doi.org/10.1016/j.tvjl.2016.04.008>

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Considerações técnicas (Jergens, 2016)

O que a análise histopatológica não é capaz ou, pouco capaz de diagnosticar?

Table 2. Gastrointestinal diseases that may not have significant histopathologic abnormalities (modified from Jergens et al., 2011)

GI diseases unaccompanied by significant histopathologic abnormalities
Motility disturbances
Brush border defects
Antimicrobial-responsive enteropathy
Secretory diarrheas
Adverse food reactions
Mucosal permeability defects
GI, gastrointestinal

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Considerações técnicas (Jergens, 2016)

Biópsia *full-thickness* x endoscópica

- *Full-thickness*
 - Transmural
 - Procedimento invasivo
 - Maior tempo de anestesia
 - Hospitalização prolongada
 - Custos
 - Inabilidade de observar lesões mucosas
 - Em gatos: indicada nas doenças do trato gastrointestinal e pancreato-biliar

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Considerações técnicas (Jergens, 2016)

Biópsia *full-thickness* x endoscópica

- Endoscópica
 - Menor tempo de procedimento
 - Uso em pacientes críticos
 - Demanda *expertise*
 - Maior chance de sub-amostragem
 - Dificuldade em obter amostras do íleo e jejuno inferior

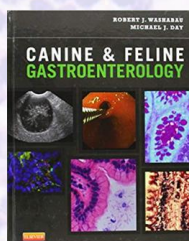
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Considerações técnicas (Jergens, 2016)

Biópsia *full-thickness* x endoscópica

- Linfoma de mucosa e transmural
- Linfoma ileal
- Linfoma e IBD concomitantes em diferentes segmentos intestinais

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CHAPTER 29

Histopathology

CHAPTER 27

Endoscopy

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Indicações e biópsia

- Esofagoscopia
 - Esofagites não granulomatosas
 - Esofagites granulomatosa (*S. lupi*)
 - Tumores esofágicos

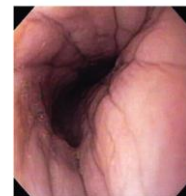
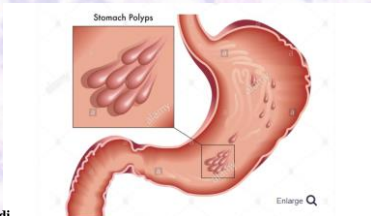


Figure 27-13 Proximal esophagus, dog. Endoscopic image of the proximal esophagus of a dog. The lumen is collapsed and longitudinal folds are visible. Air insufflation will expand the visible lumen and allow the endoscope to be safely advanced while the mucosa is examined.

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Indicações e biópsia

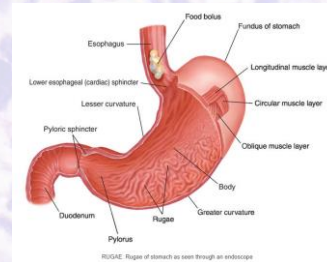
- Gastroscoopia
- Gastrites *latu sensu* (ulcerativas, erosivas, não ulcerativas e não erosivas)
- Tumores gástricos
 - Carcinomas (focal tumor like ou difusos), pólipos



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Indicações e biópsia

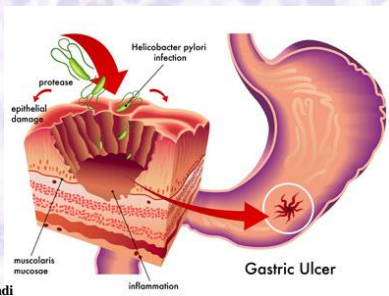
- Gastroscoopia
 - Lesão x normalidade
 - Biópsia das rugas/dobras gástricas do corpo (normalidade)



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Indicações e biópsia

- Gastroscoopia
- Gastrites erosivas, erodo-ulcerativas ou ulcerativas



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Indicações e biópsia

- Duodenoscopia, jejunoscoopia e ileoscopia
 - Diarréia crônica ou recorrente
 - Vômitos
 - Dor abdominal recorrente
 - Perda de peso
 - Hematoquezia ou melena
 - Anemias

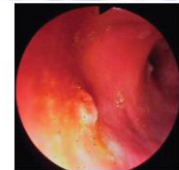


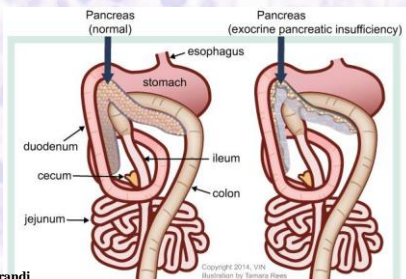
Figure 27-28 Endoscopic picture of normal small intestinal mucosa in the duodenum of a healthy patient, showing also the rugae, minor, and accessory papillae.¹¹

- Ausência de resposta ao tratamento clínico

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Indicações e biópsia

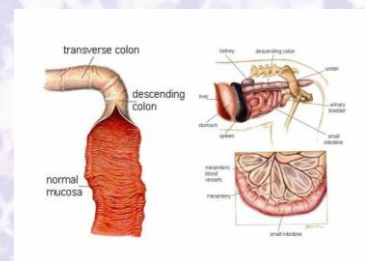
- Duodenoscopia, jejunoscoopia e ileoscopia
 - Porções passíveis de avaliação: duodeno e jejuno proximal; íleo terminal (colonoscopia)>decisão por laparoscopia



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Indicações e biópsia

- Colonoscopia
 - Análise do esfíncter fleo-cólico, ceco, cólon ascendente, transverso, descendente e canal ano-retal.
 - Íleo distal



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Indicações e biópsia

- Colonoscopia
- Tenesmo
- Disquezia
- Muco fecal
- Hematoquezia
- Hipoalbuminemia
- Perda de peso
- Anemia
- Vômitos (30% dos casos originários nas colonopatias, principalmente em gatos)

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Manipulação pós-coleta

- Técnica *free floating*

Luis Apestequí, Luis Javier Pina
Ultrasound-guided core-needle biopsy of breast lesions.



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Manipulação pós-coleta

- Técnica da esponja sintética



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Manipulação pós-coleta

- Técnica das fatias de pepino



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Manipulação pós-coleta



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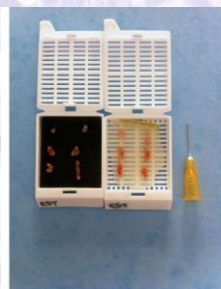


Fig 1. Gastric endoscopic biopsies following mounting and orientation on the synthetic foam sponge (left cassette) and on the thin cucumber slice (right cassette). Each line of 2 biopsy specimens corresponds to 1 site of the stomach (antrum, body, and fundus from the top to the bottom, respectively).

Manipulação pós-coleta

Journal of Veterinary Internal Medicine

ACVIM

Open Access

J Vet Intern Med 2016;30:1014-1021

Comparison of 3 Handling Techniques for Endoscopically Obtained Gastric and Duodenal Biopsy Specimens: A Prospective Study in Dogs and Cats

G.C. Ruiz, E. Reyes-Gomez, E.J. Hall, and V. Freiche

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Manipulação pós-coleta

Background: Limited evidence exists in the literature regarding whether a specific mount is preferable to use for processing endoscopically obtained gastrointestinal biopsy specimens.

Hypothesis/Objectives: To compare 3 methods of handling endoscopically obtained gastrointestinal biopsy specimens from collection to laboratory processing and to determine if any technique produced superior results.

Animals: Twenty-three dogs and cats presented for gastrointestinal signs.

Methods: Prospective study of dogs and cats presented with gastrointestinal signs to a veterinary teaching referral hospital which underwent upper gastrointestinal endoscopy. Biopsy specimens were taken from the stomach and duodenum and submitted to the laboratory using 3 techniques: mounted on a cucumber slice, mounted on a moisturized synthetic foam sponge, and floating free in formalin. The techniques were compared with regard to the specimens' width, orientation, presence of artifacts, and pathologist's confidence in diagnosis.

Results: Twenty-three patients were included, with a total of 528 biopsies collected. Specimens on cucumber slice and on sponge were significantly wider ($P < .001$ and $P = .001$, respectively) compared to those floating free in formalin (mean width of 3.81 versus 3.31 and 2.52 mm, respectively). However, specimens on synthetic sponge had significantly fewer artifacts compared to those on cucumber slice ($P = .05$) and those floating free in formalin ($P = .02$). Confidence in the diagnosis also was superior with the sponge technique over floating free specimens ($P = .002$).

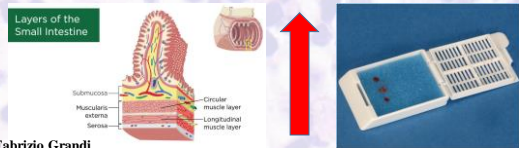
Conclusions and Clinical Importance: The use of mounted gastrointestinal biopsy specimens was superior over the use of specimens floating free in formalin. This technique improved the quality of the specimens and the pathologist's confidence in their histopathologic interpretation.

Key words: Duodenoscopy; Gastroenterology; Histopathology; Mount.

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Fixação e manuseio das amostras

- Pós-coleta
- Orientação dos fragmentos sobre fatia de pepino ou esponja sintética em colunas com as vilosidades deitadas para cima
- Fixação imediata em formol à 10%
- **Manuseio das amostras: somente após a fixação em formol à 10%**



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Fixação e manuseio das amostras



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Requisição de exame ao patologista

ENDOSCOPIC EXAMINATION REPORT: UPPER GI ENDOSCOPY

Date of procedure: _____ Case Number: _____

Patient and client information:
(card or stamp)

PROCEDURE(S): _____
Indication(s) for procedure: _____
Endoscope(s) used: _____
Forceps/retrieval device(s) used: _____

PROBLEMS/COMPLICATIONS: None
Perforation Excessive bleeding Anesthetic complications Excessive time Other

Comments: _____
 Unable to complete full examination: why? _____
 Unable to obtain adequate biopsies: why? _____
 Unable to retrieve foreign object: why? _____
 Visualization obscured: why? _____

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Requisição de exame ao patologista

SAMPLING: Biopsy Brush cytology Washing Aspiration Foreign body retrieved

DOCUMENTATION: Video Photographs

ESOPHAGUS Normal Foreign body Mass Stricture Hiatal hernia

Lesion	Code	Comments (include location)
Hyperemia/vascularity		
Discoloration		
Friability		
Hemorrhage		
Erosion/ulcer		
Contents (mucus/bile/food)		
Dilation		
Gastroesophageal sphincter		
Other		

Code: Normal = 0 Mild = 1 Moderate = 2 Severe = 3

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Requisição de exame ao patologista

STOMACH Normal Foreign body Mass Polyp(s) Parasite(s)

Site(s) of lesions: Fundus Body Incisura Antrum Pylorus

Site(s) of biopsies: Fundus Body Incisura Antrum Pylorus

Lesion	Code	Comments (include location)
Can't inflate lumen		
Hyperemia/vascularity		
Edema		
Discoloration		
Friability		
Hemorrhage		
Erosion/ulcer		
Contents (mucus/bile/food)		
Gastroesophageal sphincter		
Passing scope through pylorus		
Other		

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Requisição de exame ao patologista

DOBENUM/JEJENUM Normal Foreign body Mass Polyp Parasite(s)
 How far was the tip of the scope advanced?
 Was/were the papillae seen? Yes (which? _____) No

Lesion	Code	Comments (include location)
Can't inflate lumen		
Hyperemia/vascularity		
Edema		
Discoloration		
Friability		
Texture		
Hemorrhage		
Erosion/ulcer		
Lacteal dilatation		
Contents (mucus/bile/food)		
Other		

Code: Normal = 0 Mild = 1 Moderate = 2 Severe = 3

Comments and Recommendations: _____

Endoscopist signature: _____



This standard form was developed by the WSAVA Gastrointestinal Standardization Group (Drs Washabau, Willard, Hall, Jergens, Day, Mansell, Wilcox, Minami, Guilford, and Biltzer) with sponsorship from Hill's Pet Nutrition

Critérios de coleta e qualidade das amostras Estômago e duodeno

J Vet Intern Med 2008;22:1084-1089

Effect of Sample Quality on the Sensitivity of Endoscopic Biopsy for Detecting Gastric and Duodenal Lesions in Dogs and Cats

M.D. Willard, J. Mansell, G.T. Fosgate, M. Gualtieri, D. Olivero, P. Lecoindre, D.C. Twedt, M.G. Collett, M.J. Day, E.J. Hall, A.E. Jergens, J.W. Simpson, R.W. Else, and R.J. Washabau

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Critérios de coleta e qualidade das amostras Duodeno

• Amostra adequada

"Adequate" was defined as tissue that had at least 3 villi and had subvillus lamina propria that extended to the mucosa-muscularis mucosa border, whether or not it included muscularis mucosa.

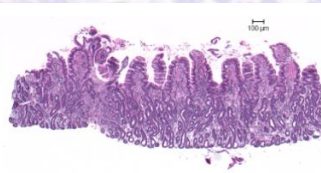


Fig 3. Photomicrograph of a biopsy sample of canine duodenum. This is an example of an "adequate" tissue sample that has at least 3 villi and encompasses the entire depth of the intestinal mucosa as seen by subvillus lamina propria, which extends to the mucosa-muscularis mucosa border. Even though muscularis mucosa is not present, the smooth, uniform lower border of the tissue sample shows that it extends to the muscularis mucosa. Hematoxylin-eosin staining.

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Critérios de coleta e qualidade das amostras Estômago e duodeno

• Amostra inadequada

"Inadequate" was defined as tissue that had only villi or subvillus lamina propria, but not both

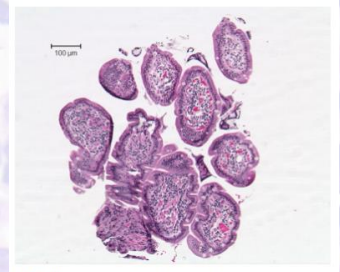


Fig 1. Photomicrograph of a biopsy sample of canine duodenum. Only villus tips are present. This is considered an "inadequate" tissue sample. Hematoxylin-eosin staining.

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Critérios de coleta e qualidade das amostras Estômago e duodeno

• Amostra marginal

"Marginal" was defined as samples that had at least 1 villus plus subvillus lamina propria, but did not clearly have the full thickness of the subvillus lamina propria extending to the muscularis mucosa

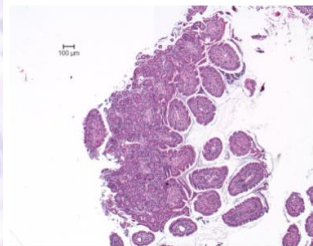


Fig 2. Photomicrograph of a biopsy sample of canine duodenum. This is an example of a "marginal" tissue sample. Hematoxylin-eosin staining.

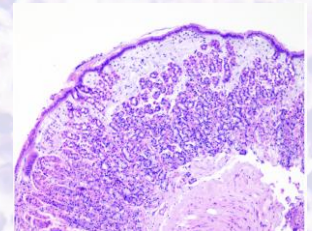
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Critérios de coleta e qualidade das amostras Estômago

"Adequate" was defined as samples that had full-thickness mucosa, whether or not it included muscularis mucosa

"Inadequate" was defined as samples that had only superficial mucosa and epithelium, or deep mucosa, but not both.

"Marginal" was defined as samples that had epithelium and mucosa, but did not clearly have full-thickness mucosa.



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Critérios de coleta e qualidade das amostras

Estômago

Table 3. Sensitivity and number of pieces of tissue required for 80-99% confidence in diagnosis of select canine and feline gastric lesions, based on the quality of the tissue samples on the histopathology slide.

Lesion	Tissue Quality	Number of Tissue Samples*	Sensitivity (%)	Number of Samples Required for Detection of Lesion			Rows Significantly Different
				80% Confidence	90% Confidence	99% Confidence	
Mild infiltrates (dogs)	Inadequate	21/7	33.3	4	6	12	a, b
	Marginal	97/73	75.3	2	2	4	a
Moderate infiltrates (dogs)	Inadequate	8/1	12.5	13	18	35	c
	Marginal	42/13	31.0	5	7	13	d
Mild infiltrates (cats)	Inadequate	212/106	50.0	3	4	7	c, d
	Marginal	22/5	22.7	7	9	18	e, f
Moderate infiltrates (cats)	Inadequate	70/52	74.3	2	2	4	e
	Marginal	177/137	77.4	2	2	4	f
Crypt lesions	Inadequate	8/0	0	ND	ND	ND	
	Marginal	54/35	64.8	2	3	5	
Moderate cellular infiltrates	Inadequate	103/60	58.3	2	3	6	
	Marginal						

ND, could not be diagnosed with tissue samples of this quality.
 *This column represents the total number of tissue samples (positive or negative) from animals with the lesion/total number of tissue samples from animals that were positive for the lesion.
 Rows with the same letter are different at a, P < .001; b, P < .001; c, P = .0037; d, P = .034; e, P < .001; f, P < .001.

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Critérios de coleta e qualidade das amostras

Duodeno

Table 1. Sensitivity and number of pieces of tissue required for 80-99% confidence in diagnosis of select duodenal lesions in dogs, based on the quality of the tissue samples on the histopathology slide.

Lesion	Tissue Quality	Number of Tissue Samples*	Sensitivity (%)	Number of Samples Required for Detection of Lesion			Rows Significantly Different
				80% Confidence	90% Confidence	99% Confidence	
Blunt villi	Inadequate	22/0	0	ND	ND	ND	
	Marginal	60/16	26.7	6	8	15	a
Lymphangiectasia	Inadequate	108/67	62.0	2	3	5	a
	Marginal	29/3	10.3	15	22	43	b,c
Crypt lesions	Inadequate	117/50	42.7	3	5	9	b
	Marginal	151/85	56.3	2	3	6	c
Mild cellular infiltrates	Inadequate	21/1	4.8	33	48	95	d
	Marginal	79/12	15.2	10	14	28	e
Moderate cellular infiltrates	Inadequate	67/15	22.4	7	10	19	e, f
	Marginal	235/118	49.4	3	4	7	e, g
Moderate cellular infiltrates	Inadequate	318/228	71.7	2	2	4	f, g
	Marginal	164/69	42.1	3	5	9	h, i
Crypt lesions	Inadequate	240/152	63.3	2	3	5	i, j
	Marginal						

ND, could not be diagnosed with tissue samples of this quality.
 *This column represents the total number of tissue samples (positive or negative) from animals with the lesion/total number of tissue samples from animals that were positive for the lesion.
 Rows with the same letter are different at a, P < .001; b, P < .001; c, P < .001; d, P < .001; e, P < .001; f, P < .001; g, P = .0037; h, P < .001; i, P = .010; j, P < .001.

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Critérios de coleta e qualidade das amostras

Duodeno

Biopsy Quality Affects Diagnosis

1087

Table 2. Sensitivity and number of pieces of tissue required for 80-99% confidence in diagnosis of select duodenal lesions in cats, based on the quality of the tissue samples on the histopathology slide.

Lesion	Tissue Quality	Number of Tissue Samples*	Sensitivity (%)	Number of Samples Required for Detection of Lesion			Rows Significantly Different
				80% Confidence	90% Confidence	99% Confidence	
Blunt villi	Inadequate	24/4	16.7	9	13	26	a, b
	Marginal	30/17	56.7	2	3	6	a
Mild cellular infiltrates	Inadequate	89/60	67.4	2	3	5	b
	Marginal	44/24	54.5	3	3	6	c
Moderate cellular infiltrates	Inadequate	81/64	79.0	2	2	3	d
	Marginal	141/136	96.5	1	1	2	c, d
Crypt lesions	Inadequate	27/20	74.1	2	2	4	
	Marginal	56/44	78.6	2	2	3	
Crypt lesions	Inadequate	109/98	89.9	1	2	3	
	Marginal						

*This column represents the total number of tissue samples (positive or negative) from animals with the lesion/total number of tissue samples from animals that were positive for the lesion.
 Rows with the same letter are different at a, P = .002; b, P < .001; c, P < .001; d, P = .008.

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Critérios de coleta e qualidade das amostras

Número mínimo de amostras EDA (Willard, 2008)

- **Gato**
 - Estômago e duodeno
 - 6 adequados ou marginais (99% de confiança de encontrar qualquer lesão)
- **Cão**
 - Duodeno
 - 6 adequados, exceto para lesões criptais ou;
 - 10-15 marginais, exceto para lesões criptais.
 - Lesões criptais
 - 13 adequados ou >20 marginais
 - Estômago
 - 7 adequados ou 13 marginais

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Critérios de coleta e qualidade das amostras

Exemplos de amostragem

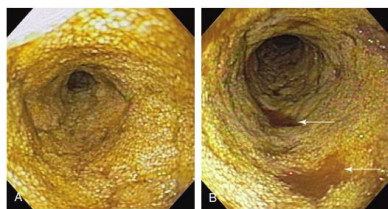


Figure 17-40. A, Endoscopic view of the duodenal mucosa of a Pug with a severe protein-losing enteropathy (serum albumin 1.4 g/dL). The mucosa appears uniformly normal throughout this image. B, Endoscopic view of the same duodenum. This image was taken several centimeters further aboral. Now focal areas that appear to have loss of villi (i.e., the surface is depressed relative to the rest of the mucosa and is smooth, arrows) can be seen. However, there is still more normal-appearing mucosa than abnormal-appearing mucosa. C, Photomicrograph of a tissue sample from the depressed, smooth areas seen in (B). Note that there is an intact epithelial surface but a complete loss of villi plus a marked inflammatory cell infiltrate. D, Endoscopic view of the duodenum of a Beagle Terrier with severe diarrhea and protein-losing enteropathy. Note that there are areas of the mucosa that are relatively light colored (small arrows) and other areas that are darker red in color (larger arrows). Also note that the darker red areas comprise a much smaller portion of the surface area of the duodenal mucosa. E, Photomicrograph of the light-colored duodenal mucosa seen in (D). Note that this is relatively normal except for some villus blunting. F, Photomicrograph of the darker-colored duodenal mucosa seen in (D). Note that there is fusion of villi (despite an intact but damaged and attenuated epithelial surface) and severe distortion of mucosal crypts. This is severe mucosal disease.

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Critérios de coleta e qualidade das amostras

Exemplos de amostragem

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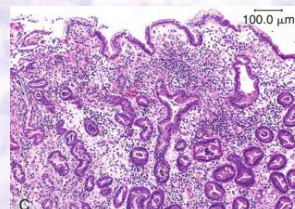


Figure 17-40. A, Endoscopic view of the duodenal mucosa of a Pug with a severe protein-losing enteropathy (serum albumin 1.4 g/dL). The mucosa appears uniformly normal throughout this image. B, Endoscopic view of the same duodenum. This image was taken several centimeters further aboral. Now focal areas that appear to have loss of villi (i.e., the surface is depressed relative to the rest of the mucosa and is smooth, arrows) can be seen. However, there is still more normal-appearing mucosa than abnormal-appearing mucosa. C, Photomicrograph of a tissue sample from the depressed, smooth areas seen in (B). Note that there is an intact epithelial surface but a complete loss of villi plus a marked inflammatory cell infiltrate. D, Endoscopic view of the duodenum of a Beagle Terrier with severe diarrhea and protein-losing enteropathy. Note that there are areas of the mucosa that are relatively light colored (small arrows) and other areas that are darker red in color (larger arrows). Also note that the darker red areas comprise a much smaller portion of the surface area of the duodenal mucosa. E, Photomicrograph of the light-colored duodenal mucosa seen in (D). Note that this is relatively normal except for some villus blunting. F, Photomicrograph of the darker-colored duodenal mucosa seen in (D). Note that there is fusion of villi (despite an intact but damaged and attenuated epithelial surface) and severe distortion of mucosal crypts. This is severe mucosal disease.

Técnicas de coloração

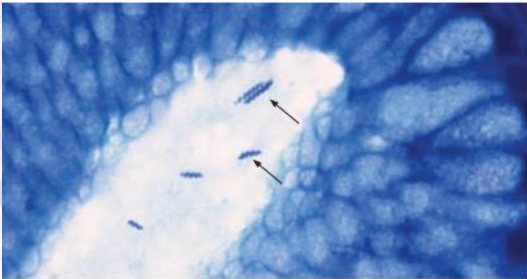
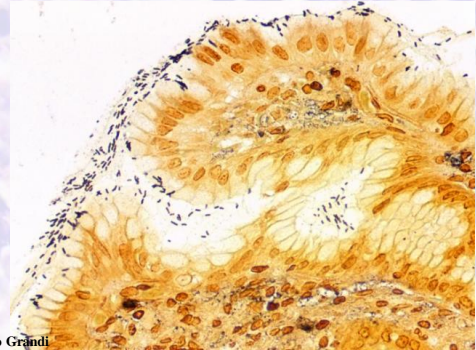


Figure 1 Giemsa stain shows *Helicobacter heilmannii* organisms ($\times 100$).

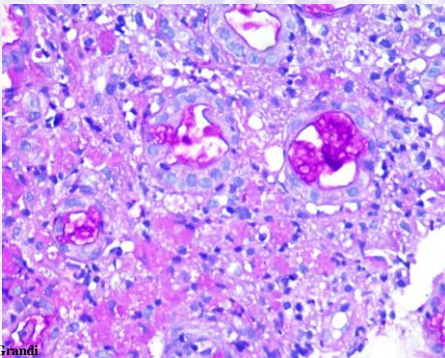
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Técnicas de coloração



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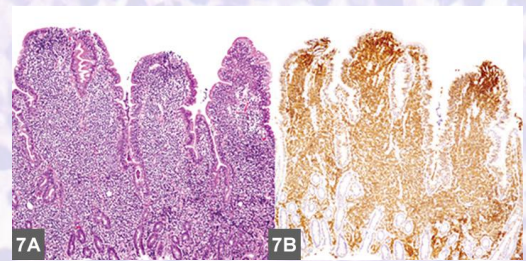
Técnicas de coloração



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Imuno-histoquímica

Fonte: clinician's brief

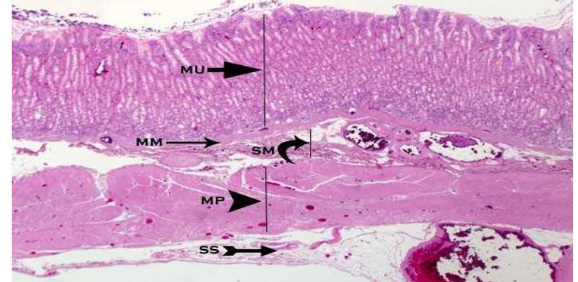


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Histologia e histopatologia gástrica:
padrões de resposta inflamatória

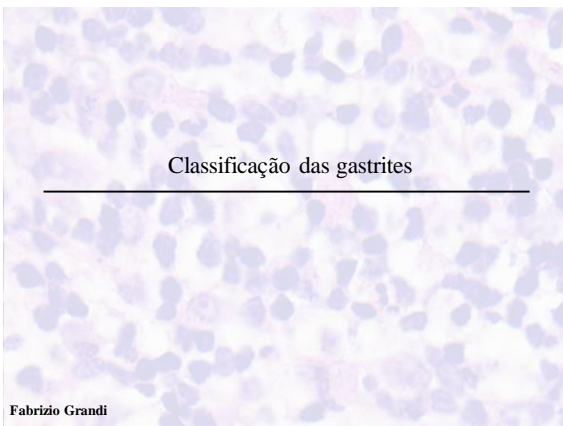
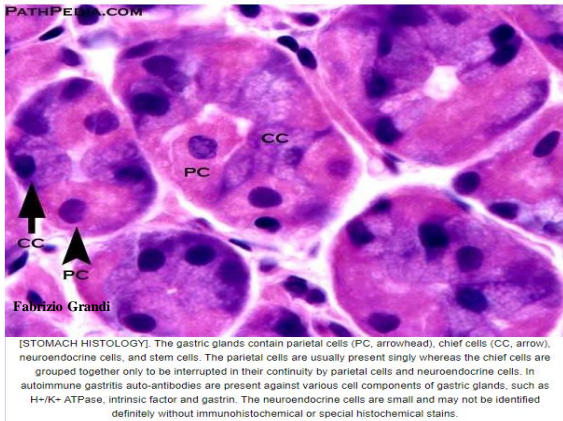
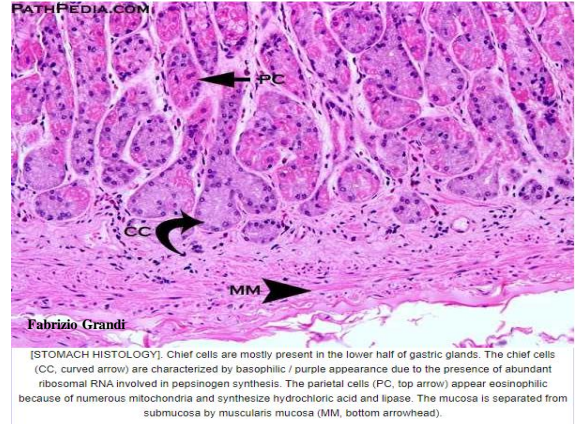
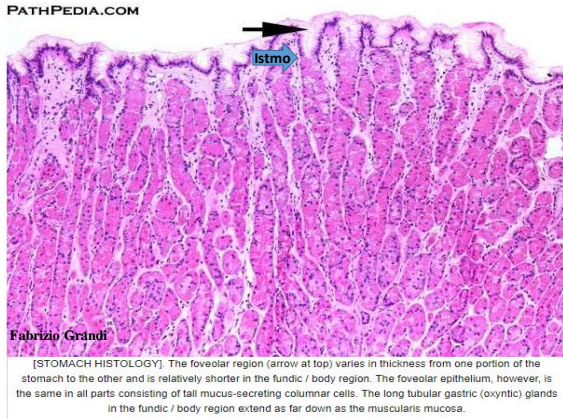
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[STOMACH HISTOLOGY]. Stomach, like other segments of the gastrointestinal tract, consists of mucosa (MU, top arrow), submucosa (SM, curved arrow), muscularis propria (MP, arrowhead), and serosa (SS, arrow with tail). The mucosa consists of superficial foveolar epithelium and deeper glands. The cardiac and antral regions consist of mucus-secreting glands whereas the fundic / body region consists of parietal and chief cells. The mucosa is separated from submucosa by a thin muscularis mucosa (MM, thin middle arrow).




J. Comp. Path. 2008, Vol. 138, 81-843 Available online at www.sciencedirect.com
 ScienceDirect ELSEVIER
Histopathological Standards for the Diagnosis of Gastrointestinal Inflammation in Endoscopic Biopsy Samples from the Dog and Cat: A Report from the World Small Animal Veterinary Association Gastrointestinal Standardization Group
 M. J. Day*, T. Bilzer[†], J. Mansell[‡], B. Wilcock[§], E. J. Hall[¶], A. Jergens^{||}, T. Minami[¶], M. Willard[¶] and R. Washabau[¶]
 * University of Bristol, Bristol, UK; [†] University of Düsseldorf, Düsseldorf, Germany; [‡] Texas A&M University, College Station, TX, USA; [§] Histvet, Guelph, Canada; [¶] Iowa State University, Ames, IA, USA; [¶] Pet-Vet, Yokohama, Japan and [¶] University of Minnesota, St Paul, MN, USA
 Fabrizio Grandi

INFLAMMATION				
Intraepithelial lymphocytes	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria lymphocytes and plasma cells	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria eosinophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria neutrophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Other inflammatory cells	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Gastric lymphofollicular hyperplasia	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
FINAL DIAGNOSIS				
Normal tissue	<input type="checkbox"/>			
Lymphoplasmacytic inflammatory	<input type="checkbox"/>			
Eosinophilic inflammatory	<input type="checkbox"/>			
Neutrophilic inflammatory	<input type="checkbox"/>			
Mucosal atrophy/fibrosis (non-inflammatory)	<input type="checkbox"/>			
Other	<input type="checkbox"/>			
OTHER COMMENTS				
Fabrizio Grandi				

INFLAMMATION				
Intraepithelial lymphocytes	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria lymphocytes and plasma cells	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria eosinophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria neutrophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Other	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
FINAL DIAGNOSIS				
Normal tissue	<input type="checkbox"/>			
Lymphoplasmacytic inflammatory	<input type="checkbox"/>			
Eosinophilic inflammatory	<input type="checkbox"/>			
Neutrophilic inflammatory	<input type="checkbox"/>			
Lymphangiectasia	<input type="checkbox"/>			
Mucosal atrophy/fibrosis (non-inflammatory)	<input type="checkbox"/>			
Other	<input type="checkbox"/>			
OTHER COMMENTS				
Fabrizio Grandi				

Gastrointestinal Histopathology Standards


Fabrizio Grandi  **IWSAVA**
 Global Veterinary Development

STANDARD FORM FOR ASSESSMENT OF THE GASTRIC BODY OR ANTRAL MUCOSA

Pathologist _____ Case number _____
 Number of pieces of gastric tissue on slide _____
 Tissue present
 Inadequate Too superficial Adequate depth
 Number of tissues abnormal _____

MORPHOLOGICAL FEATURES

	Normal	Mild	Moderate	Marked
Surface epithelial injury	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Gastric pit epithelial injury	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Fibrosis/glandular nesting/mucosal atrophy	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

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STANDARD FORM FOR ASSESSMENT OF DUODENAL MUCOSA

Pathologist _____ Case number _____
 Number of pieces of duodenal tissue on slide _____
 Tissue present
 Inadequate Too superficial Adequate depth
 Number of tissues abnormal _____

MORPHOLOGICAL FEATURES

	Normal	Mild	Moderate	Marked
Villous stunting	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Epithelial injury	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Crypt distension	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lacteal dilation	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Mucosal fibrosis	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Gastrointestinal Histopathology Standards

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STANDARD FORM FOR ASSESSMENT OF COLONIC MUCOSA

Pathologist _____ Case number _____
 Number of pieces of colonic tissue on slide _____
 Tissue present
 Inadequate Too superficial Adequate depth
 Number of colonic tissues abnormal _____

MORPHOLOGICAL FEATURES

	Normal	Mild	Moderate	Marked
Surface epithelial injury	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Crypt hyperplasia	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Crypt dilation/distortion	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Fibrosis/atrophy	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

INFLAMMATION

Lamina propria lymphocytes and plasma cells	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria eosinophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria neutrophils	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Lamina propria macrophages	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

FINAL DIAGNOSIS

Normal colon	<input type="checkbox"/>
Lymphoplasmacytic inflammatory	<input type="checkbox"/>
Eosinophilic inflammatory	<input type="checkbox"/>
Neutrophilic inflammatory	<input type="checkbox"/>
Histiocytic/granulomatous inflammatory	<input type="checkbox"/>
Mucosal atrophy/fibrosis (non-inflammatory)	<input type="checkbox"/>
Other	<input type="checkbox"/>

OTHER COMMENTS

Fabrizio Grandi



Original Article

Correlating Gastrointestinal Histopathologic Changes to Clinical Disease Activity in Dogs With Idiopathic Inflammatory Bowel Disease

Karin A. Allenspach¹, Jonathan P. Mochel¹, Yingzhou Du¹, Simon L. Priestnall², Frances Moore³, Michael Slayter⁴, Aline Rodrigues⁵, Mark Ackermann⁶, Mark Krockenberger⁶, Joanne Mansell⁷, WSAVA GI Standardization Working Group⁸, Nicole Luckschander⁷, Chong Wang¹, Jan Suchodolski¹, Nora Berghoff⁹, and Albert E. Jergens¹

Veterinary Pathology
 JLR
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 DOI: 10.1177/0898010118813000
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Fabrizio Grandi



Abstract

Prior studies have failed to detect a convincing association between histologic lesions of inflammation and clinical activity in dogs with inflammatory bowel disease (IBD). We hypothesized that use of a simplified histopathologic scoring system would improve the consistency of interpretation among pathologists when describing histologic lesions of gastrointestinal inflammation. Our aim was to evaluate the correlation of histopathologic changes to clinical activity in dogs with IBD using this new system. Forty-two dogs with IBD and 19 healthy control dogs were enrolled in this retrospective study. Endoscopic biopsies from the stomach, duodenum, ileum, and colon were independently scored by 8 pathologists. Clinical disease activity was scored using the Canine Inflammatory Bowel Disease Activity Index (CIBDAI) or the Canine Chronic Enteropathy Clinical Activity Index (CCECAI), depending on the individual study center. Summative histopathological scores and clinical activity were calculated for each tissue (stomach, duodenum, ileum, and colon) and each tissue histologic score (inflammatory/morphologic feature). The correlation between CCECAI/CIBDAI and summative histopathologic score was significant ($P < .05$) for duodenum ($r = 0.42$) and colon ($r = 0.33$). In evaluating the relationship between histopathologic scores and clinical activity, significant ($P < .05$) correlations were observed for crypt dilation ($r = 0.42$), lamina propria (LP) lymphocytes ($r = 0.40$), LP neutrophils ($r = 0.45$), mucosal fibrosis ($r = 0.47$), lacteal dilation ($r = 0.39$), and villus stunting ($r = 0.43$). Compared to earlier grading schemes, the simplified scoring system shows improved utility in correlating histopathologic features (both summative histology scores and select histologic scores) to IBD clinical activity.

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Allenspach et al

7

Table 5. Quantitative Simplified Scoring System for Defining Gastrointestinal Inflammation.

Location	Histopathologic Parameter	Grade			
		0 (Normal)	1 (Mild)	2 (Moderate)	3 (Marked)
Stomach (fundus)	Morphologic parameter				
	Fibrosis (number of fibrocytes separating glands)	≤2	3–5	6–10	≥11
	Intraepithelial lymphocytes (lymphocytes per stretch of 50 epithelial cells)	≤2	3–10	11–20	≥21
	Lamina propria lymphocytes and plasma cells (cells per 400 × field)	≤20	21–50	51–100	≥101
Inflammatory parameters	Lamina propria eosinophils (cells per 400 × field)	≤2	3–20	21–50	≥51
	Lamina propria neutrophils (cells per 400 × field)	0	≤20	21–50	≥51
Stomach (antrum)	Morphologic parameter				
	Fibrosis (number of fibrocytes separating gastric pits or mucous glands)	≤10	11–15	16–20	≥21
Inflammatory parameters	Intraepithelial lymphocytes (lymphocytes per stretch of 50 epithelial cells)	≤2	3–5	4–10	≥11
	Lamina propria lymphocytes and plasma cells (cells per 400 × field)			As fundus	
	Lamina propria eosinophils (cells per 400 × field)	≤2	3–10	11–50	≥51
	Lamina propria neutrophils (cells per 400 × field)			As fundus	

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Gastrite superficial de mucosa

- Critérios diagnósticos
 - Infiltração de 20% da lâmina própria superficial
 - Edema
 - Ativação de fibroblastos e endotélio
 - Linfocítica
 - Plasmocitária
 - Linfoplasmocitária (maioria)
 - Eosinofílica

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Gastrite superficial de mucosa

- Possibilidades diagnósticas
 - Gastrite superficial de mucosa, linfoplasmocitária, severa.
 - Gastrite superficial de mucosa, linfoplasmocitária
 - Gastrite superficial de mucosa, linfoplasmocitária e eosinofílica, moderada
 - Gastrite superficial de mucosa (itens marcados a parte)
 - Gastrite por *Helicobacter* spp (helicobacteriose)
 - Gastrite superficial de mucosa, linfoplasmocitária, severa por *Helicobacter* spp.
 - Gastrite superficial de mucosa, linfoplasmocitária, severa. Negativo ou positivo para *Helicobacter* spp.
 - Gastrite crônica, superficial de mucosa, linfoplasmocitária, severa. Negativa ou positiva para *Helicobacter* spp.

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Gastrite transmucosa

- Critérios diagnósticos
 - Lâmina própria *full-thickness*
 - Fibrose
 - Edema
 - Nidação e atrofia glandular
 - Infiltrado inflamatório
 - Linfocítica
 - Plasmocitária
 - Linfoplasmocitária (maioria)
 - Eosinofílica

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Gastrite atrófica

- Critérios diagnósticos
 - Entidade pouco reconhecida em cães e gatos (lesão residual da gastrite transmucosa)
 - Inflamação crônica
 - Atrofia glandular
 - Fibrose e nidação
 - Regeneração glandular
 - Hiperplasia linfocitocitária
 - Redução do número de células parietais
 - Aumento do número de células produtoras de muco

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Gastrite atrófica

- Critérios diagnósticos
 - Norwegian Lundehund (associação entre carcinoma gástricos e gastrite atrófica)



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Gastrite ulcerativa

- Critérios diagnósticos
 - Basofilia e *flattening* do epitélio superficial
 - Perda das células produtoras de muco do istmo
 - Expansão de células germinativas no istmo
 - Infiltrado inflamatório mínimo
 - Neutrófilos
 - Causas
 - Abrasão mecânica
 - Estimulo histamínico (*MCT*)
 - Ingestão de produtos químicos
 - Drogas (AINES e AIES)
 - Exercício (cães de corrida)

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Gastropatia urêmica

- Critérios diagnósticos
 - Mineralização e necrose das células parietais no terço médio da mucosa gástrica
 - Mineralização da MB glandular e dos vasos
 - Mineralização do musculo liso da MM ou MP

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Gastropatia por vasculite

- CID em cães
- PIF em gatos

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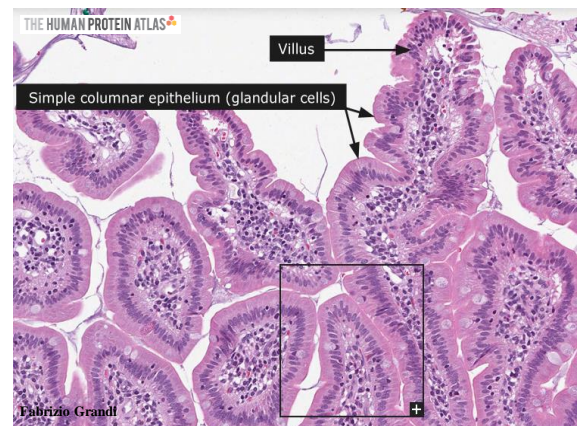
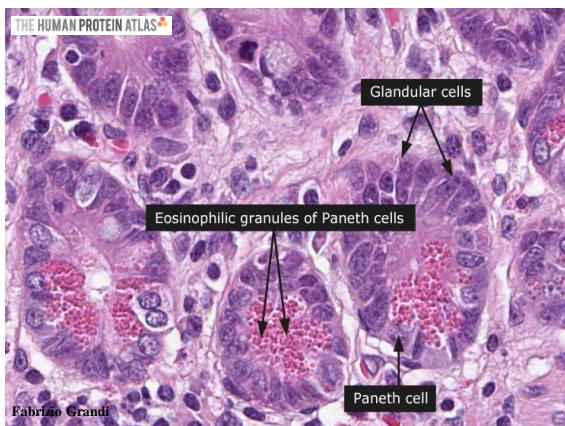
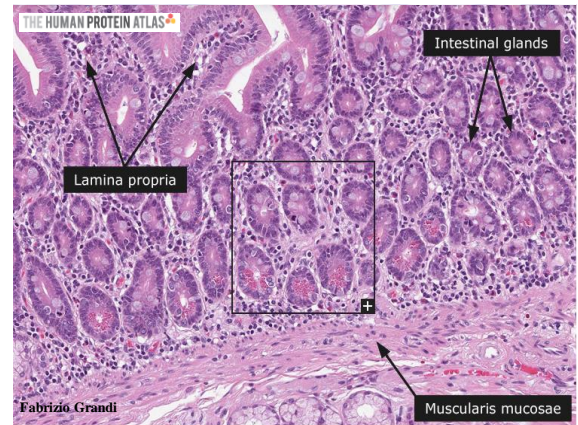
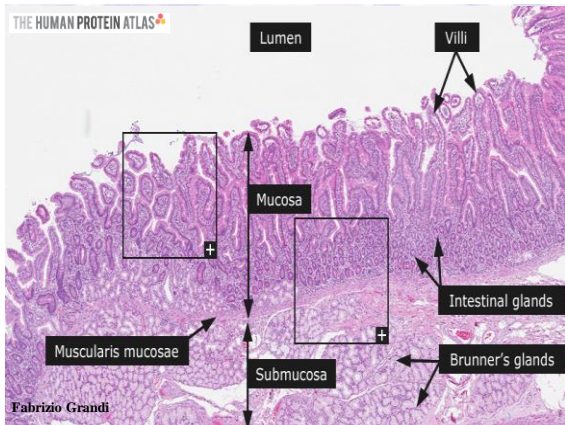
Outros gastrites

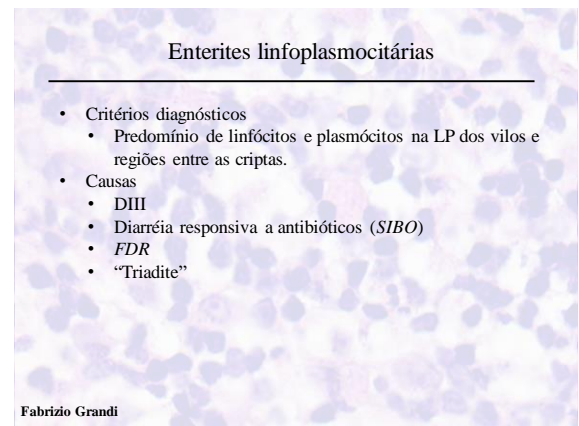
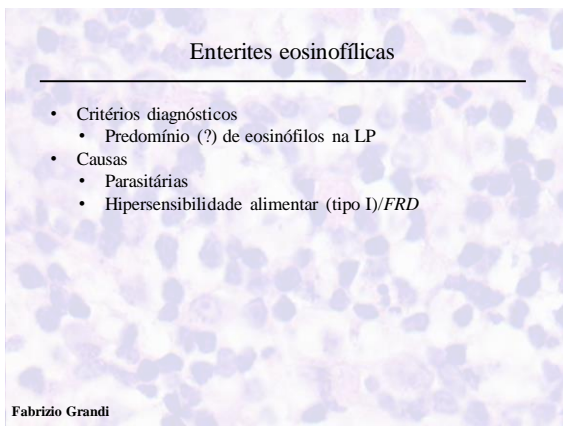
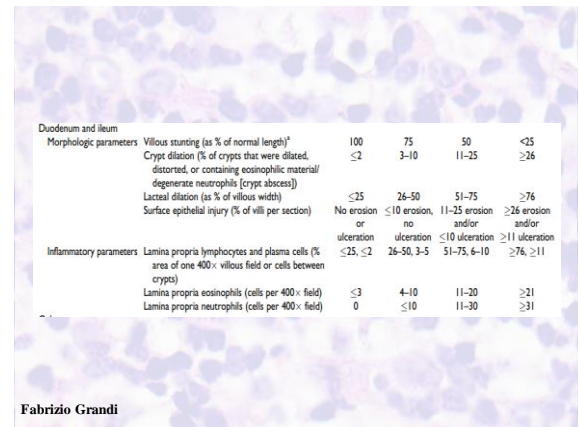
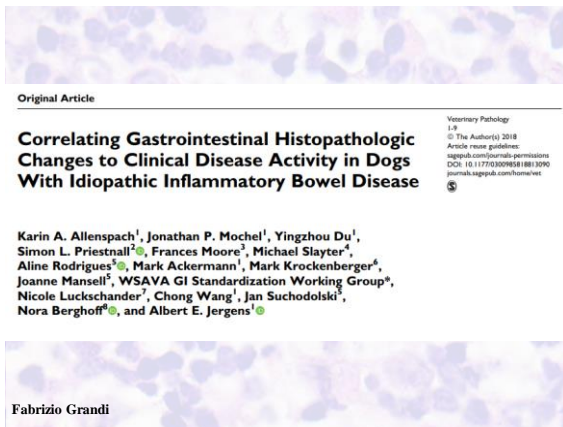
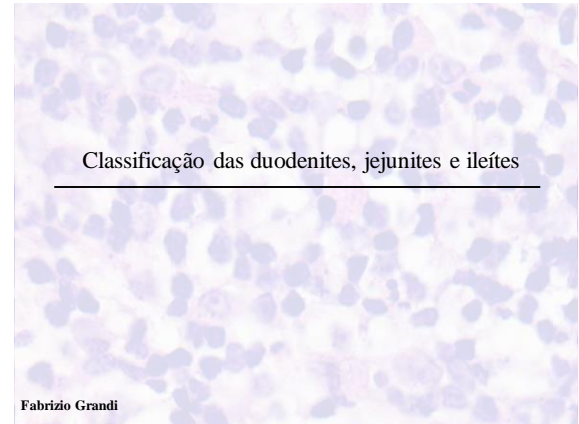
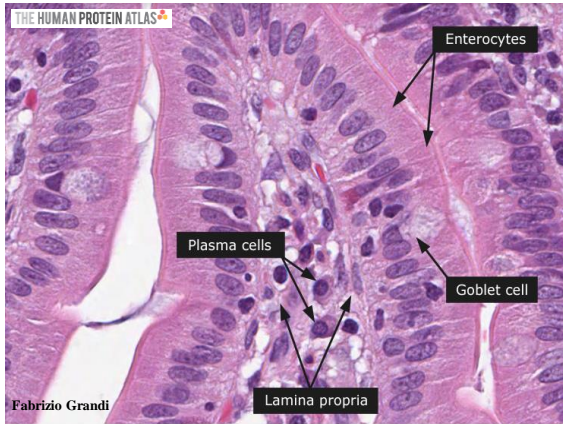
- Parasitárias: *Physaloptera spp* (cães), *Ollulanus spp* (gatos), *Gnathostoma spp*, *Cylicospirura spp*
- *P. insidiosum*
- *H. pylori*

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Histologia e histopatologia do intestino delgado:
padrões de resposta inflamatória

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Enterites neutrofilicas

- Critérios diagnósticos
 - Predomínio (?) de neutrófilos na LP
- Causas
 - Infeciosas

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Enterites hemorrágicas ou necro-hemorrágicas

- Critérios diagnósticos
 - Necrose das vilosidades (pontas ou completa)
 - Necrose e regeneração das criptas
 - Encurtamento e fusão de vilosidades
 - Hemorragia em LP
 - Fibrina na superfície dos enterócitos
 - Neutrófilos em LP
 - Bacilos gram + aderidos aos enterócitos e restos necróticos



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Enterites associadas a raça

Tissue Antigens

Genetic susceptibility to gluten sensitive enteropathy in Irish setter dogs is not linked to the major histocompatibility complex

A. Polvi, O.A. Garden, R.S. Houlston, M. Maki, R.M. Batt, J. Partanen

First published: 11 December 2008 | <https://doi.org/10.1111/j.1399-0039.1998.tb03085.x> | Cited by: 26

J Vet Intern Med 2000;14:69-80

Familial Protein-Losing Enteropathy and Protein-Losing Nephropathy in Soft Coated Wheaten Terriers: 222 Cases (1983-1997)

Meryl P. Littman, Donna M. Dambach, Shelly L. Vaden, and Urs Giger

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Enterites associadas a raça

Vet. Pathol. 25:36-41 (1988)

Gastroenteritis of Basenji Dogs

N. J. MACLACHLAN, E. B. BREITSCHWERDT, J. M. CHAMBERS, R. A. ARGENZIO, AND E. V. DE BUYSCHER

Departments of Microbiology, Pathology, and Parasitology; Companion Animals and Special Species Medicine, and Anatomy, Physiology, and Radiology, School of Veterinary Medicine, North Carolina State University, Raleigh, NC, and The University of North Carolina Core Center in Diarrheal Diseases, Chapel Hill, NC



Protein-losing enteropathy in the Lunde hund

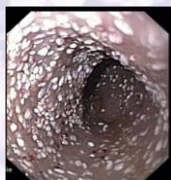
Kjell Fiesjö, Torstein Yvi

Fabrizio Grandi

First published: January 1977 | <https://doi.org/10.1111/j.1748-5827.1977.tb05819.x> | Cited by: 28

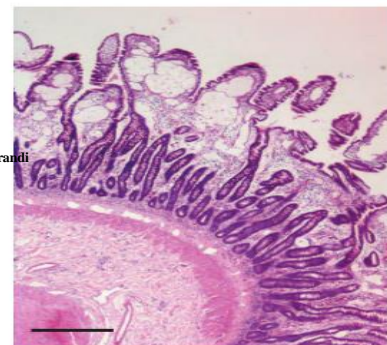
Linfangiectasia

- Critérios diagnósticos
 - Dilatação dos vasos linfáticos da LP das vilosidades ou entre as criptas
 - Edema
 - Encurtamento das vilosidades
 - Vasos linfáticos da submucosa, muscular, serosa e mesentérico podem ser acometidos
 - Primária (congenita) ou secundária



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Fonte: Wikivet



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Figure 29-22 Lymphangiectasia. In this full-thickness biopsy there is marked ballooning dilation of the villus lacteals with reduction in the height of the affected villi. There is also lymphatic dilation within the pericryptal mucosa and the submucosa. Hematoxylin and eosin stain; bar = 1 mm.

Linfangite lipogranulomatosa intestinal

- Critérios diagnósticos
 - Dilatação de linfáticos
 - Fibrose
 - Macrófagos epitelióides ou xantomatosos
 - Células gigantes multinucleadas
 - Infiltrado linfoplasmocitário e neutrofílico
 - Cristais de colesterol aciculares

J Vet Intern Med 2014;28:48–51

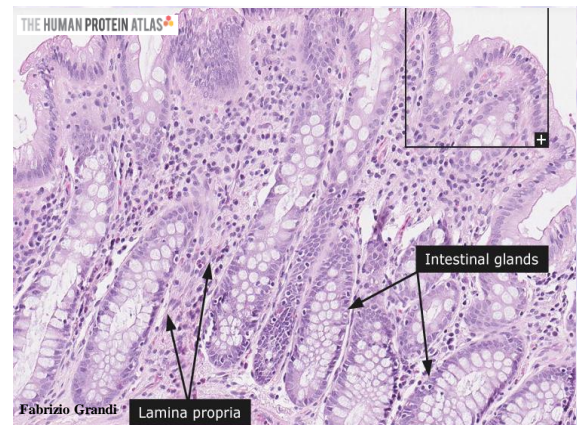
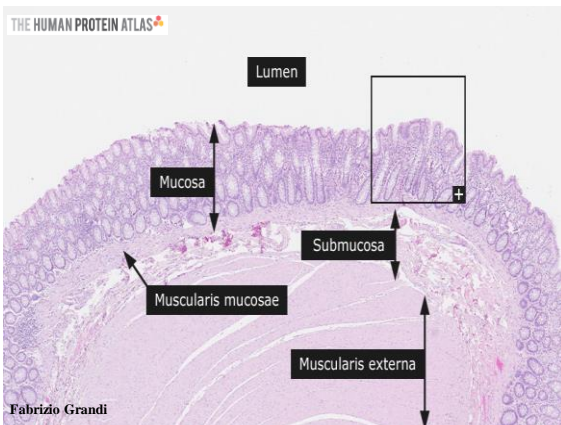
Focal Intestinal Lipogranulomatous Lymphangitis in 6 Dogs (2008–2011)

V.E. Watson, M.M. Hobday, and A.C. Durham

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Histologia e histopatologia do cólon: padrões de resposta inflamatória

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Classificação das colites

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Box 29-1 The Fundamental Rules of Colonic Biopsy Interpretation

- Colonic inflammatory disease is virtually always diffuse and uniform, so numerous biopsies are unnecessary.
 - Because even very small decreases in absorptive efficiency can result in profound diarrhea, even very subtle colonic lesions can be clinically significant.
 - Because colonic ulceration heals very rapidly and usually without any residual changes, biopsies must be taken during active clinical disease so as to avoid the risk of a misleading false-negative result.
 - Leukocytes should not exceed four cell layers between adjacent crypts, and eosinophils should not appear in the superficial half of the mucosa.
 - Credible etiologic candidates are rarely seen.
- Fabrizio Grandi

Original Article

Correlating Gastrointestinal Histopathologic Changes to Clinical Disease Activity in Dogs With Idiopathic Inflammatory Bowel Disease

Veterinary Pathology
1-9
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DOI: 10.1177/0899765518810200
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Karin A. Allenspach¹, Jonathan P. Mochel¹, Yingzhou Du¹, Simon L. Priestnall², Frances Moore¹, Michael Slayter¹, Aline Rodrigues³, Mark Ackermann¹, Mark Krockenberger⁴, Joanne Mansell⁵, WSAVA GI Standardization Working Group⁶, Nicole Luckschander⁷, Chong Wang¹, Jan Suchodolski¹, Nora Berghoff⁸, and Albert E. Jergens⁹

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Colon

Morphologic parameters	Crypt dilation and distension (% of crypts per section)	0	≤25	26-50	≥51
	Fibrosis (number of fibrocytes separating crypts)	≤2	3-5	6-10	≥11
	Goblet cell numbers (% reduction from normal)	0	≤25	26-50	≥51
	Surface epithelial injury (% of villi per section)		As duodenum and ileum		
Inflammatory parameters	Lamina propria lymphocytes and plasma cells (cells between crypts)	≤5	6-10	11-20	≥21
	Lamina propria eosinophils (cells per 400x field)	≤2	3-10	11-20	≥21
	Lamina propria neutrophils (cells per 400x field)		As duodenum and ileum		
	Lamina propria macrophages (cells per 400x field)	≤2	3-20	21-50	≥51

Fabrizio Grandi

Colite linfoplasmocitária (e eosinofílica) idiopática de mucosa

- 90% dos casos em cães e gatos
- Critérios diagnósticos
 - Edema
 - Fibrose
 - > 4 colunas de plasmócitos e linfócitos entre as criptas (Brian Wilcock)
 - Ulceração presente ou ausente

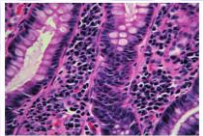


Figure 25-38. Transverse section of mucosal colitis with an increase in mucosal lymphoplasmocytic infiltrate between adjacent crypts, mucosal edema, and crypt fibrosis. This is by far the most common pattern of colitis in dogs and cats, with no proven etiologic implications. Hematoxylin and eosin stain.

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Colite eosinofílica

- Critérios diagnósticos
 - Predomínio (?) de eosinófilos na LP
 - Presença de eosinófilos na metade superior da lâmina própria

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Colite granulomatosa de mucosa

- Critérios diagnósticos
 - Predomínio de macrófagos na LP
 - Ex. histoplasmose e colite ulcerativa histiocítica

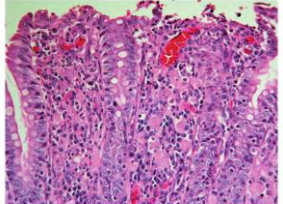


Figure 25-39. Early histiocytic ulcerative colitis in a young Boxer dog. The early lesion is usually found deep in the mucosa, associated with epithelial hyperplasia and flattening as evidence for ongoing superficial ulceration. Hematoxylin and eosin stain.

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Colite ulcerativa histiocítica

- Bulldog Francês e Boxers
- Critérios diagnósticos
 - Predomínio de macrófagos PAS positivos na LP
 - Erodo-ulcerações colônicas

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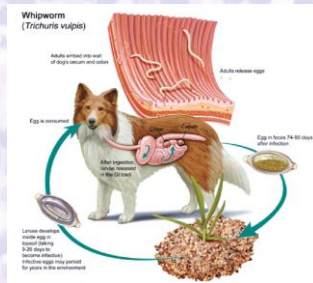


Journal of Comparative Pathology
Volume 122, Issues 2-3, February 2000, Pages 163-175

Regular Article
An Immunohistochemical Study of Histiocytic Ulcerative Colitis in Boxer Dogs
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Colite ulcerativa

- Etiologia
 - T. foetus* em gatos
 - T. vulpis* em cães



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Colite de submucosa e transmural

- Padrão morfológico com alta importância diagnóstica
- Etiologia
 - PIF
 - Crítérios diagnósticos
 - Vasculite necrotizante de submucosa
 - Edema e hemorragia
 - Necrose vascular
 - Mucosa normal
 - Neutrofílica, granulomatosa ou piogranulomatosa
 - P. insidiosum*

Fabrizio Grandi